The effect of prostaglandin E_1 on the increase of serum lactate and plasma granulocyte elastase activity during radical surgery for esophageal cancer

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Abstract: Serum lactate concentrations and the lactate/pyruvate (L/P) ratio were measured in two groups of patients undergoing radical esophagectomy, as an indicator of tissue hypoxia, and β-glucuronidase and granulocyte elastase as indicators of tissue damage. One group received prostaglandin E_1 (PGE₁) and the other group received nothing. Serum lactate concentrations and the L/P ratio increased significantly 30 min after starting thoracotomy in the patients who were not treated with PGE₁. On the contrary, intravenous drip infusion of PGE₁ (0.04 µg·kg⁻¹·min⁻¹) suppressed the increases in serum lactate concentratons and L/P ratios. Plasma granulocyte elastase activity increased significantly at the end of surgery in both groups. There was no change in serum β -glucuronidase activity in both groups. This study suggests that low doses of PGE₁ maintain organ blood flow without affecting blood pressure. However, these low doses of PGE₁ could not suppress granulocyte elastase release.

Key words: Lactate, Prostaglandin E₁, Esophageal cancer

Introduction

Postoperative pulmonary complications associated with thoracotomy and mediastinal lymph node dissection for esophageal cancer have been reported [1,2]. These complications may progress to multiple organ failure (MOF). Pathogenic factors of MOF include organ ischemia related to circulatory insufficiency and the effect of humoral mediators, indicating the importance of maintaining blood flow during surgery [3–5]. Prostaglandin E_1 (PGE₁) is known to be a vasodilator and to increase organ blood flow, and has direct cytoprotective effects such as cellular membrane stabilization [6–8].

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In this study, the effect of PGE_1 therapy was assessed by measuring serum concentrations of lactate and pyruvate as an indicator of tissue hypoxia, and β -glucuronidase and granulocyte elastase (GEL) activity as indicators of tissue damage.

Patients and methods

After institutional approval of the study protocol, informed consent was obtained from 30 patients. All underwent radical surgery for esophageal cancer. Preoperative medication consisted of atropine 0.01 mg·kg⁻¹, hydroxyzine 1 mg·kg⁻¹, and pethidine 1 mg·kg⁻¹ was administered intramuscularly 1 h before arrival in the operating room.

ECG, a pulse oximeter (Datex, IMI), and a capnometer (Datex, Capnomac 873379-15, IMI) were monitored continuously. A radial arterial catheter was inserted after induction of anesthesia and direct arterial pressure was measured. After induction of anesthesia with thiamylal ($5 \text{ mg} \cdot \text{kg}^{-1}$), the trachea was intubated with a double-lumen tube using succinylcholine chloride. Anesthesia was maintained with N₂O in 50% O₂ and isoflurane (2%) and muscle relaxation was achieved with pancuronium (0.1 mg \cdot \text{kg}^{-1}). Supplementary fluid infusion was accomplished with intravenous drip infusion of 10 ml \cdot \text{kg}^{-1} \cdot hr^{-1} Ringer's lactate solution and 5 units of fresh frozen plasma.

The patients were divided into two groups: one not receiving PGE_1 (group I) and the other receiving the drug (group II). PGE_1 (0.04 μ g·kg⁻¹·min⁻¹) was given by intravenous drip infusion throughout the surgery.

In each patient, four blood samples were drawn from the radial artery, one each at the time of inducing anesthesia, 30 and 60 min after starting thoracotomy, and at the end of the surgery. Serum lactate, pyruvate, β -glucuronidase and plasma GEL activity were measured in those blood samples.

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Activity of the complex of polymorphonuclear (PMN) elastase with α_1 -protease inhibitor (α_1 -PI) was determined by the sandwich immunoassay method [9], and the results of this assay were used as GEL activity. β -glucuronidase was assayed by enzyme immunoassay. Serum lactate and pyruvate were measured by the enzymatic method.

The data was statistically analyzed by analysis of variance and Neuman-Keuls multiple comparison tests. The level of significance was defined as P < 0.05. All statistics are given as the mean \pm standard deviation (SD).

Results

There were no significant differences with regard to the patients' age, body weight, and duration of surgery between groups I and II (Table 1).

Mean arterial pressure increased significantly at 30 min after thoracotomy compared with the preoperative level in group I. There were no other changes in either group (Table 2).

Serum lactate levels increased significantly, compared with the values before surgery, from 30 min after thoracotomy up to the end of surgery in both groups. In group II, the increase of serum lactate concentration at the end of surgery was lower than that in group 1 (P <0.01) (Table 2) (Fig. 1). In both groups, the L/P ratio increased significantly at the end of surgery (P < 0.01) (Table 2).

Plasma GEL activity increased significantly at the end of surgery compared with the value before surgery in each group (P < 0.05) (Table 2) (Fig. 2). Serum β glucuronidase activity did not change significantly in either group (Table 2).

The volume of urine output during surgery was 2.6 \pm 1.3 ml·kg⁻¹·hr⁻¹ in group II, much more than 1.5 \pm 0.7 ml·kg⁻¹·hr⁻¹ in group I (P < 0.01).

Discussion

Blood lactate concentration is a valuable indicator of the severity of surgical stress. Hyperlactacidemia appears in two situation. First, it occurs when the common metabolic pool for lactate and pyruvate increases; that is, no change in redox with a constant ratio of lactate to pyruvate. Second, it is caused by an elevated L/P ratio with a shift in redox to the direction of reduction. An elevated L/P ratio reflects a high NADH/NAD ratio, suggesting that NADH accumulates in the cytoplasm due to tissue hypoxia or impairment of the respiratory chain in mitochondria.

In this study, we observed the increase in serum lactate levels from 30 min after the start of thoracotomy during radical surgery for esophageal cancer. At the end of the surgery, the L/P ratio increased significantly,

Table 1. Patient's age, body weight, and duration of surgery (mean \pm SD)

	n	Age (years)	Body weight (kg)	Duration of surgery (min)	
Group I	15	63 ± 7.3	51 ± 5.6	373 ± 45.3	
Group II	15	66 ± 8.5	51 ± 7.5	397 ± 64.3	

Table 2. Value of serum lactate, lactate/pyruvate ratio (L/P ratio), β -glucuronidase, plasma granulocyte elastase (*GEL*) activity, and mean arterial pressure (*MAP*) in each period (mean \pm SD)

		Before surgery	30 min after thoracotomy	60 min after thoracotomy	End of surgery
Lactate	group I	11.2 ± 3	14.8 ± 3*	15.9 ± 3**	$31.8 \pm 5^{**}$
(mg/dl)	group II	12.2 ± 2	15.9 ± 2	$17.6 \pm 2*$	$25.5 \pm 6^{***}$
L/P ratio	group I	11.0 ± 1	11.9 ± 1	12.0 ± 1	$15.2 \pm 4*$
	group II	10.9 ± 1	12.1 ± 1	12.9 ± 1	$13.2 \pm 1.6^*$
GEL activity	group 1	232 ± 104	256 ± 105	271 ± 127	$386 \pm 148*$
(µg/l)	group II	160 ± 19	152 ± 24	161 ± 21	$351 \pm 106*$
β-glucuronidase	group I	556 ± 174	590 ± 194	599 ± 190	654 ± 183
(µg/dl)	group II	577 ± 102	662 ± 59	634 ± 219	747 ± 156
MAP (mmHg)	group I	80 ± 15	98 ± 7*	92 ± 9	81 ± 14
	group II	78 ± 18	92 ± 13	90 ± 14	82 ± 10

* P < 0.05 compared with value before surgery; ** P < 0.01 compared with value before surgery; * P < 0.01 compared with value in group I. **Fig. 1.** Changes in serum lactate concentration (mean \pm SD) for control (*closed circles*) and prostaglandin E₁ (PGE₁, *open circles*). **P* < 0.05 compared with value before surgery; ***P* < 0.01 compared with value before surgery; **P* < 0.01 compared with value before surgery; **P* < 0.01 compared with value in group I sampling points: 1, before surgery after induction of anesthesia; 2, 30 min after thoracotomy; 3, 60 min after thoracotomy; and 4, the end of surgery

These results suggest that prolonged massive surgical stress affects circulation in various organs due to excessive stimulation of the autonomic nerves [10], resulting in ischemia in the peripheral tissues. In hypoxia, oxidative phosphorylation of mitochondria is suppressed which leaves anaerobic glycolysis as the major source of ATP production, followed by a rise in pyruvate. In the reaction of pyruvate + NADN + H⁺ \rightarrow lactate + NAD⁺ by lactate dehydrogenase (LDH), the lactate level is much higher than pyruvate at equilibrium, causing lactate to accumulate [11,12].

Normally, lactate is taken up in mitochondria where it is oxidized after entering the citric acid cycle following reaction with pyruvate dehydrogenase (PDH). In the liver and kidneys, however, it is consumed as the substrate for glyconeogenesis. When oxygen delivery to the tissues is impaired, lactate production increases, while lactate consumption simultaneously decreases due to impaired mitochondrial function. As a result, the serum lactate concentration increases significantly [12].

With tissue hypoxia, the liver, which is primarily responsible for metabolizing lactate, begins to produce it [13]. PGE₁ decreases blood pressure by its vasodilating effect on peripheral arteries, while it is reported to maintain blood flow and thus protect the heart, kidneys, liver, and brain [14–17]. When tissue perfusion is improved by decreased peripheral vascular resistance, the production of lactate in the peripheral tissues seems to be suppressed. It has been reported that PGE_1 regulates the energy balance in hepatocytes and intracellular electrolyte balance by activating adenyl cyclase in hepatocytes, and thus by increasing cyclic AMP. Okabe et al. [18] reported that PGI_2 attenuates the reduction in oxidative phosphorylation of liver mitochondria due to ischemia [18]. In this study, the accumulation of lactate was inhibited by PGE_1 . This seemed to be due to a synergistic effect between increased hepatic and renal blood flow and improved mitochondrial function in the liver and kidneys.

 PGE_1 has been reported to increase renal plasma flow, urine output, and Na excretion [19–21]. The present results support previous findings that urine output was significantly higher in patients receiving PGE_1 than in those not receiving the drug.

Plasma GEL is one of the chemical mediators which cause postoperative pulmonary complications. In both groups, it increased significantly at the end of surgery. These results may be explained by the release of GEL following aggregation of polymorphonuclear cells associated with mechanical stimulation of the lungs during thoracotomy. GEL is one of the neutral proteases which lyse or destroy pulmonary components including collagen, elastin, and fibronectin, resulting in increased vascular permeability [22–25], and affect postoperative pulmonary complications.

Nuytinck et al. [26] studied multiple organ failure following trauma and suggested that GEL was the best



Fig. 2. The changes of plasma granulocyte elastase (*GEL*) activity (mean \pm SD) for control (*closed circles*) and PGE₁

(open circles). *P < 0.05 compared with value before surgery

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mg/dl

35

25

15

5

Serum Lactate

predictor of adult respiratory distress syndrome (ARDS) and MOF, and was related to the survival rate of those who developed these diseases. These results indicate the importance of intraoperative management to suppress the increase in GEL activity. We had expected that PGE₁ suppress GEL by stabilizing the membrane; however, it did not show this effect at the relatively low dose of $0.04 \,\mu g \cdot k g^{-1} \cdot min^{-1}$ used in this study.

 PGE_1 has been reported to maintain organ blood flow without affecting blood pressure, when given at low doses of $0.02-0.04 \,\mu g \cdot k g^{-1} \cdot min^{-1}$. The results of this study suggest that low doses of PGE_1 can be used safely without producing excessive hypotension. In conclusion, serum lactate concentration and the L/P ratio increased significantly during radical surgery for esophageal cancer. The treatment with PGE_1 inhibited the increase of serum lactate. It is suggested that PGE_1 maintains blood flow in the vital organs. However, low doses of PGE_1 could not suppress the increase in granulocyte elastase.

References

- Nakayama K, Kakegawa T (1981) Latest management of pulmonary complications following esophageal cancer surgery in Japan. Int Adv Surg Oncol 4:111–125
- Nagano M, Kawasaki J, Horiuchi T, et al. (1981) Post-operative mechanical ventilatory support and circulatory care for the esophageal cancer surgery (in Japanese). Kokyu to Junkan 29:159–164
- 3. Carrico CJ, Meakins JL, Marshall JC, et al. (1986) Multipleorgan-failure syndrome. Arch Surg 121:196-208
- Goris RJA, Boekhorst TPA, Nuytinch JKS, et al. (1985) Multipleorgan-failure. Arch Surg 120:1109–1115
- 5. Redl H, Paul E, Goris RJA, et al. (1988) Plasma levels of elastase α_1 protease inhibitor complex in the monitoring of ARDS and multi-organ failure; A summary of three clinical trials. Adv Exp Med Biol 240:457–464
- Ronald GT, Ingo M, Gerhard S, et al. (1988) Hepatic reperfusion injury following orthotopic liver transplantation in the rat. Transplantation 46:502–506
- 7. Ferguson WW, Edmonds AW, Starling JR, et al. (1973) Protective effect of prostaglandin E_1 (PGE₁) on lysosomal enzyme release in serotonin induced gastric ulceration. Ann Surg 177:648–654
- Machiedo GW, Rush BF Jr (1979) Comparison of corticosteroids and prostaglandins in treatment of hemorrhagic shock. Ann Surg 190:735-739

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- Neumann S, Gunzer G, Hennrich N, Dang H (1984) "PMNelastase assay": Enzyme immunoassay for human polymorphonuclear elastase complexed with a₁-protease inhibitor. J Clin Chem Clin Biochem 22:693–697
- Reilly J (1934) Hemorrhagies lesions vasculaires et lymphaticus du tube digestif determinées par injection perisplanchnique de substances diverses. C R Soc de Biol 116:24
- Robert AK (1980) Lactate homeostasis and lactic acidosis. Ann Intern Med 92:227-237
- Barry A (1987) Controversies in lactic acidosis. JAMA 258: 497-501
- Park R, Aroeff AI (1980) Lactic acidosis. Adv Intern Med 25:35– 39
- 14. Feldman RL, Rose B, Verbust KM (1988) Hemodynamic and angiographic effect of prostaglandin E_1 in coronary artery disease. Am J Cardiol 62:698–702
- Popat KD, Pitt B, Mich AA (1982) Hemodynamic effects of prostaglandin E₁ infusion in patients with acute myocardial infarction. Am Heart J 103:485-489
- Geumei A, Bashour FA, Swamy BV, et al. (1973) Prostaglandin E₁; Its effects on hepatic circulation in dogs. Pharmacology 9:336– 347
- Adachi H, Sugihara H, Nakagawa H, et al. (1984) Effect of prostaglandin E₁ on fractional distribution of cardiac output and organ blood flow in man; a simultaneous and noninvasine determination using double dose thallium-201 scintigraphy. Cardiovasc Res 18:657-662
- Okabe K, Msalchesky PS, Nose Y (1986) Protective effect of prostaglandin I₂ on hepatic mitochondrial function of the preserved rat liver. Tohoku J Exp Med 150:373–378
- Dunn MJ, Hood VL (1977) Prostaglandins and kidney. Am J Physiol 233:F169-184
- 20. Jonston HH, Herzog HH, Lauler DP (1967) Effect of prostaglandin E_1 on renal hemodynamics, sodium, and water excretion. Am J Physiol 213:939–946
- 21. Goto F, Otani E, kato S, et al. (1982) Prostaglandi
n $\rm E_1$ as a hypotensive drug during general anaesthesia. Anaesthesia 37:530–535
- 22. Yasutake A, Powers JC (1981) Reactivity of human leukocyte elastase and procine pancreatic elastase toward peptide 4nitroanilides containing model desmosine residues: Evidence that human leukocyte elastase is selective for crosslinked regions of elastin. Biochemistry 20:3675-3679
- Kobayashi S, Nagai Y (1978) Human leukocyte neutral protease, with special reference to collagen metabolism. J Biochem 84:559– 567
- Harlan J, Killen P, Harkec L, et al. (1981) Neutrophil-mediated endothelial injury in vitro; Mechanisms of cell detachment. J Clin Invest 68:1394–1403
- Richards PS, Saba TM, Del Vecchio PJ (1986) Matrix fibronectin disruption in association with altered endothelial cell adhesion induced by activated polymorphonuclear leukocytes. Exp Mol Pathol 45:1-21
- Nuytinck JKS, Goris RJA, Redl H, et al. (1986) Posttraumatic complications and inflammatory mediators. Arch Surg 121:886– 890